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(FILE 'HOME' ENTERED AT 13:55:22 ON 17 JUL 2002)

INDEX 'ADISALERTS, ADISINSIGHT, ADISNEWS, AGRICOLA, ANABSTR, AQUASCI,  
BIOBUSINESS, BIOCOMMERCE, BIOSIS, BIOTECHABS, BIOTECHDS, BIOTECHNO,  
CABA,  
CANCERLIT, CAPLUS, CEABA-VTB, CEN, CIN, CONFSCI, CROPB, CROPUS, DDFB,  
DDFU, DGENE, DRUGB, DRUGLAUNCH, DRUGMONOG2, ...' ENTERED AT 13:55:35 ON  
17 JUL 2002

SEA (PHOSPHOLIPASE C)

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73 FILE ADISALERTS  
11 FILE ADISINSIGHT  
461 FILE AGRICOLA  
82 FILE ANABSTR  
147 FILE AQUASCI  
97 FILE BIOBUSINESS  
25 FILE BIOCOMMERCE  
16923 FILE BIOSIS  
152 FILE BIOTECHABS  
152 FILE BIOTECHDS  
6502 FILE BIOTECHNO  
950 FILE CABA  
3056 FILE CANCERLIT  
15686 FILE CAPLUS  
26 FILE CEABA-VTB  
7 FILE CEN  
4 FILE CIN  
348 FILE CONFSCI  
3 FILE CROPB  
26 FILE CROPUS  
153 FILE DDFB  
825 FILE DDFU  
601 FILE DGENE  
153 FILE DRUGB  
5 FILE DRUGNL  
990 FILE DRUGU  
7 FILE DRUGUPDATES  
112 FILE EMBAL  
13290 FILE EMBASE  
5582 FILE ESBIOSBASE  
366 FILE FEDRIP  
57 FILE FROSTI  
136 FILE FSTA  
1778 FILE GENBANK  
2 FILE HEALSAFE  
104 FILE IFIPAT  
1173 FILE JICST-EPLUS  
7 FILE KOSMET  
4846 FILE LIFESCI  
4 FILE MEDICONF  
14192 FILE MEDLINE  
32 FILE NIOSHTIC  
70 FILE NTIS  
11 FILE OCEAN  
58 FILE PASCAL  
12 FILE PHAR

12 FILE PHIN  
26 FILE ROMT  
15886 FILE SCISEARCH  
7208 FILE TOXCENTER  
1900 FILE USPATFULL  
15 FILE USPAT2  
201 FILE WPIDS  
201 FILE WPINDEX  
L1 QUE (PHOSPHOLIPASE C)  
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FILE 'AGRICOLA, BIOSIS, CAPLUS, SCISEARCH, EMBASE, MEDLINE, BIOTECHNO'  
ENTERED AT 13:57:24 ON 17 JUL 2002  
L2 5 S L1 AND (ANIMAL(W) FEED)  
L3 0 S L2 AND (CEREUS)  
L4 4 DUP REM L2 (1 DUPLICATE REMOVED)  
L5 2596 S L1 AND COMPOSITION  
L6 1086 S L5 AND PHOSPHATIDYLINOSITOL  
L7 54 S L6 AND (CEREUS)  
L8 33 DUP REM L7 (21 DUPLICATES REMOVED)

L8 ANSWER 23 OF 33 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.  
ACCESSION NUMBER: 1986:148960 BIOSIS  
DOCUMENT NUMBER: BA81:59376  
TITLE: INFLUENCE OF PHOSPHOLIPID METABOLITES ON THE FUSION OF MODEL MEMBRANES OF DIFFERENT COMPOSITION.  
AUTHOR(S): SHRAGIN A S; VASILENKO I A; SELISHCHEVA A A; SHVETS V I  
CORPORATE SOURCE: M.V. LOMONOSOV MOSC. INST. FINE CHEM. TECHNOL., MOSCOW, USSR.  
SOURCE: BIOL MEMBR, (1985) 2 (8), 789-794.  
CODEN: BIMEE9.  
FILE SEGMENT: BA; OLD  
LANGUAGE: Russian  
AB The fusion of monolayer liposomes induced by **phospholipases C** and **D** was studied using <sup>31</sup>P-NMR spectroscopy and fluorescence of **Tb**<sup>3+</sup> complexes. **Phospholipase C** induced fusion could be observed in all cases, independently of composition of liposomes. **Phospholipase D** evoked fusion of liposomes if the latter contained a high percentage of phospholipids not prone to bilayer formation. The **phosphatidylinositol** metabolites (1,2-diacylglycerol and phosphatidic acid), formed under the action of **phospholipases C** and **D**, apparently facilitate generation of a metastable state in the membrane that in turn might perturb the bilayer structure of membrane lipids and induce their fusion.

L8 ANSWER 24 OF 33 CAPLUS COPYRIGHT 2002 ACS  
ACCESSION NUMBER: 1985:181317 CAPLUS  
DOCUMENT NUMBER: 102:181317  
TITLE: Electrophoretic characterization of hepatic alkaline phosphatase released by **phosphatidylinositol**-specific **phospholipase C**. A comparison with liver membrane and serum-soluble forms  
AUTHOR(S): Kominami, Tatsuya; Miki, Akira; Ikehara, Yukio  
CORPORATE SOURCE: Sch. Med., Fukuoka Univ., Fukuoka, 814-01, Japan  
SOURCE: Biochem. J. (1985), 227(1), 183-9  
CODEN: BIJOAK; ISSN: 0306-3275  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB Alk. phosphatase (I) was solubilized from plasma membrane of rat liver with BuOH, bile acids or Na deoxycholate, and electrophoretically compared with a sol. form in blood serum which was derived from the liver. Three I preps. from the plasma membrane migrated at the same position on polyacrylamide-gel electrophoresis (PAGE) in the presence of either Triton X-100 or SDS. Their mobility, however, was distinctly different from that of the serum-sol. form of liver-derived I. On the other hand, **phosphatidylinositol**-specific **phospholipase C** isolated from *Bacillus cereus* was used to release I from plasma membrane. Released I had the same mobility as the serum-sol. form on PAGE in the presence or absence of detergents. **Phospholipase C** also converted the BuOH-extd. membrane form into the serum-sol. form. Apparently, release of I from the liver into serum is not simply caused by a detergent effect of bile salts, but involves an enzymic hydrolysis of **phosphatidylinositol**, with which I may strongly interact in the membrane.

ACCESSION NUMBER: 1983:301584 BIOSIS  
DOCUMENT NUMBER: BA76:59076  
TITLE: ASYMMETRY OF LIPID ORGANIZATION IN CHOLINERGIC SYNAPTIC VESICLE MEMBRANES.  
AUTHOR(S): MICHAELSON D M; BARKAI G; BARENHOLZ Y  
CORPORATE SOURCE: LAB. NEUROCHEM., DEP. BIOCHEM., HEBREW UNIV.-HADASSAH MED. SCH., JERUSALEM 91010, ISRAEL.  
SOURCE: BIOCHEM J, (1983) 211 (1), 155-162.  
CODEN: BIJOAK. ISSN: 0306-3275.  
FILE SEGMENT: BA; OLD  
LANGUAGE: English

AB The lipid **composition** of purified *Torpedo ocellata* cholinergic synaptic vesicles was determined, and their distribution between the inner

and outer leaflets of the vesicular membrane was investigated. The vesicles contain cholesterol and phospholipids at a molar ratio of 0.63. The vesicular phospholipids are as follows (mol% of total phospholipids): phosphatidylcholine (40.9); phosphatidylethanolamine (24.6); plasmenylethanolamine (11.5); sphingomyelin (12); phosphatidylserine (7.3); **phosphatidylinositol** (3.7). The asymmetry of the synaptic vesicle membranes was investigated by 2 independent approaches: determining accessibility of the amino lipids to the chemical label trinitrobenzenesulfonic acid (TNBS), and determining accessibility of the vesicular glycerophospholipids to **phospholipase C** (*Bacillus cereus*). TNBS rendered the vesicles leaky and cannot be used reliably to determine the asymmetry of *T. ocellata* synaptic vesicle membranes. Incubation of the vesicles with **phospholipase C** (*B. cereus*) results in biphasic hydrolysis of the vesicular glycerophospholipids. About 45% of the phospholipids are hydrolyzed in < 1 min, during which no vesicular acetylcholine is released. In the 2nd phase, the hydrolysis of the phospholipids slows down

markedly and is accompanied by loss of all the vesicular acetylcholine. The lipids hydrolyzed during the 1st phase were those comprising the outer

leaflet. Analysis of the results thus obtained indicates that the vesicular membrane is asymmetric: all the **phosphatidylinositol**, 77% of the phosphatidylethanolamine, 47% of the plasmenylethanolamine and 58% of the phosphatidylcholine resided in the outer leaflet. Since phosphatidylserine is a poor substrate for **phospholipase C** (*B. cereus*), its distribution between the 2 leaflets of the synaptic vesicle membrane is only suggestive.

ACCESSION NUMBER: 1982:522809 CAPLUS  
DOCUMENT NUMBER: 97:122809  
TITLE: Study of the lipid dependence of pyrophosphatase activity in microsomes from rat liver and hepatoma by the use of **phospholipase C**  
AUTHOR(S): Dyatlovitskaya, E. V.; Yaronskaya, E. B.; Bergel'son, L. D.  
CORPORATE SOURCE: M. M. Shemyakin Inst. Bioorg. Chem., Moscow, USSR  
SOURCE: Biokhimiya (Moscow) (1982), 47(7), 1222-9  
CODEN: BIOHAO; ISSN: 0006-307X  
DOCUMENT TYPE: Journal  
LANGUAGE: Russian  
AB The lipid dependence of pyrophosphatase activity was studied by treatment of liver and hepatoma microsomes with **phospholipase C** from *Clostridium perfringens* and *Bacillus cereus* and a subsequent incorporation of various classes of phospholipids into the delipidated microsomes. **Phospholipase C** hydrolysis sharply lowers the pyrophosphatase activity of liver and hepatoma

microsomes. Enzyme activity is restored after introduction of phospholipids into delipidated liver microsomes, the maximal effect being achieved on incorporation of phosphatidylcholine. All the phospholipids tested exerted the same reactivation effects on the delipidated microsomes of hepatoma. However, a more complete delipidation of hepatoma microsomes by **phospholipase C** hydrolysis and subsequent org. solvent extrn. revealed a specific dependence of the enzyme activity on phosphatidylserine.

L8 ANSWER 27 OF 33 CAPLUS COPYRIGHT 2002 ACS

ACCESSION NUMBER: 1982:610874 CAPLUS

DOCUMENT NUMBER: 97:210874

TITLE: In vitro effect of **phospholipase C** from *Bacillus cereus* on tissue

thromboplastin from different species

AUTHOR(S): Hetland, O.; Janson, T. L.; Johnsen, B.

CORPORATE SOURCE: Res. Inst. Intern. Med., Univ. Oslo, Oslo, Norway

SOURCE: Thromb. Res. (1982), 28(1), 93-101

CODEN: THBRAA; ISSN: 0049-3848

DOCUMENT TYPE: Journal

LANGUAGE: English

AB Purified **phospholipase C** from *B. cereus*

caused a significant loss in the procoagulant activity of thromboplastin preps. from man, rabbit, sheep, cow, rat, and mouse. However, marked differences were obsd. with respect to the degree of inactivation. Rat, mouse, bovine, and 1 type of rabbit preps. (prepd. from Me<sub>2</sub>CO-powd. brain) were markedly more sensitive to attack by **phospholipase C** than were preps. of human, sheep, and std. rabbit preps. The relative amts. of the individual phospholipids in thromboplastin preps. showed only minor variations among the species. The effect of **phospholipase C** on each of these phospholipids in the various thromboplastin preps. showed some significant differences.

L8 ANSWER 28 OF 33 CAPLUS COPYRIGHT 2002 ACS

ACCESSION NUMBER: 1982:2835 CAPLUS

DOCUMENT NUMBER: 96:2835

TITLE: Studies on potassium ion-proton ATPase. IV. Effects of **phospholipase C** treatment

AUTHOR(S): Schrijen, J. J.; Omachi, A.; Van Groningen-Luyben, W. A. H. M.; De Pont, J. J. H. M.; Bonting, S. L.

CORPORATE SOURCE: Dep. Biochem., Univ. Nijmegen, Nijmegen, 6500 HB, Neth.

SOURCE: Biochim. Biophys. Acta (1981), 649(1), 1-12

CODEN: BBACAO; ISSN: 0006-3002

DOCUMENT TYPE: Journal

LANGUAGE: English

AB The total phospholipid content of a gradient-purified (K<sup>+</sup> + H<sup>+</sup>)-ATPase prepn. from pig gastric mucosa is 105 .mu.mol/100 mg protein and consists of 29% sphingomyelin, 29% phosphatidylcholine, 28% phosphatidylethanolamine, 10% phosphatidylserine, and 4%

**phosphatidylinositol**. The cholesterol content corresponds to 50 .mu.mol/100 mg protein. Treatment with **phospholipase C**

(I) (from *Clostridium welchii* and *Bacillus cereus*) results in an immediate decrease of the phosphate content. Up to 50% of the phospholipids are hydrolyzed by each I prepn. alone, without further hydrolysis by increased I concn. or prolonged incubation time. Combined treatment with the 2 I preps., sequentially or simultaneously,

hydrolyzes

.1toreq.65% of the phospholipids. The (K<sup>+</sup> + H<sup>+</sup>)-ATPase and K<sup>+</sup>-stimulated p-nitrophenylphosphatase activities are decreased proportionally with the total phospholipid content, indicating that these enzyme activities are dependent on phospholipids. I treatment does not change optimal pH, Km value for ATP, and temp. dependence of the gastric (K<sup>+</sup> + H<sup>+</sup>)-ATPase, but slightly decreases the Ka value for K<sup>+</sup>. I treatment loweres adenylyl

5'-imidodiphosphate binding and phosphorylation capacities, suggesting that inactivation occurs primarily on the substrate binding level. Most of the results can be understood by assuming that hydrolysis of the phospholipids by I leads to aggregation of the membrane protein mols. and complete inactivation of the aggregated ATPase mols.

L8 ANSWER 29 OF 33 CAPLUS COPYRIGHT 2002 ACS  
ACCESSION NUMBER: 1980:492618 CAPLUS  
DOCUMENT NUMBER: 93:92618  
TITLE: Saline washing and the erythrocyte membrane  
AUTHOR(S): Rumsby, M. G.; Little, C.; White, M.; Tovey, L. A. D.  
CORPORATE SOURCE: Dep. Biol., Univ. York, Heslington/York, YO1 5DD, Engl.  
SOURCE: J. Appl. Biochem. (1979), 1(5-6), 430-41  
CODEN: JABIDV; ISSN: 0161-7354  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB Fresh human erythrocytes given a single saline wash and then resuspended in platelet- and leukocyte-free plasma contg. acid-citrate-dextrose anticoagulant released Hb more rapidly upon storage at 4.degree. than did controls. The single saline wash converted .apprx.70% of the fresh discocytes into the echinocyte I form, a process which was not reversed when the cells were resuspended and stored in their own plasma at 4.degree.. Repeated saline washing with the glass effect eliminated converted .apprx.75% of fresh erythrocytes into echinocytes I and II with practically no echinocyte III formation. Repeated saline washing of erythrocytes from 8-wk-old blood, contg. echinocytes III and spherocytocytes I but free of discocytes had no effect on the morphol. compn. The total lipid content in suspensions of fresh discoid erythrocytes decreased on repeated saline washing from 39.9 to 29.6 .times. 10-11 .mu.mol phospholipid/cell and from 32.6 to 27.3 .times. 10-11 .mu.mol cholesterol/cell. High concns. of **phospholipase C** (*Bacillus cereus*) caused no significant lysis of fresh discoid erythrocytes either in whole blood or after repeated washing. In vitro-aged echinocyte III and spherocytocyte I cell forms were much more susceptible to **phospholipase C**-induced lysis, and washing these cell forms in saline made them more sensitive to lysis though their shape was not altered. Apparently, the echinocyte I and II forms induced by saline washing in the absence of the glass effect are due to the removal of plasma factors from the outer leaflet of the red cell membrane surface, and more profound changes in membrane structure are needed to induce red cell transformation to spheroidal forms.

L8 ANSWER 30 OF 33 CAPLUS COPYRIGHT 2002 ACS  
ACCESSION NUMBER: 1978:419039 CAPLUS  
DOCUMENT NUMBER: 89:19039  
TITLE: **Phosphatidylinositol** as the endogenous activator of the (sodium-potassium ion)-dependent ATPase in microsomes of rabbit kidney  
AUTHOR(S): Mandersloot, J. G.; Roelofsen, B.; De Gier, J.  
CORPORATE SOURCE: Lab. Biochem., Univ. Utrecht, Utrecht, Neth.  
SOURCE: Biochim. Biophys. Acta (1978), 508(3), 478-85  
CODEN: BBACAQ; ISSN: 0006-3002  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB Incubation of rabbit kidney microsomes with pig pancreatic phospholipase A2 produced residual membrane preps. with very low (Na<sup>+</sup> + K<sup>+</sup>)-ATPase activity. The activity was restored by recombination with lipid vesicles of neg.-charged glycerophospholipids. Vesicles of pure phosphatidylcholine and phosphatidylethanolamine were virtually inactive in this respect, but did reactivate in the presence of cholate. Incubation of the microsomes with a combination of **phospholipase C** (*Bacillus cereus*) and sphingomyelinase C (*Staphylococcus aureus*) resulted in 90-5% release of the phospholipids.

The residual membrane contained only **phosphatidylinositol** and still showed 50-1 of the (Na<sup>+</sup> + K<sup>+</sup>)-ATPase activity.

L8 ANSWER 31 OF 33 CAPLUS COPYRIGHT 2002 ACS  
ACCESSION NUMBER: 1978:559392 CAPLUS  
DOCUMENT NUMBER: 89:159392  
TITLE: Studies on **phosphatidylinositol**  
phosphodiesterase (**phospholipase C**  
type) of *Bacillus cereus*. II. In vivo and  
immunochemical studies of phosphatase-releasing  
activity  
AUTHOR(S): Ohyabu, Tetsuo; Taguchi, Ryo; Ikezawa, Hiroh  
CORPORATE SOURCE: Fac. Pharm. Sci., Nagoya City Univ., Nagoya, Japan  
SOURCE: Arch. Biochem. Biophys. (1978), 190(1), 1-7  
CODEN: ABBIA4; ISSN: 0003-9861  
DOCUMENT TYPE: Journal  
LANGUAGE: English

AB A **phosphatidylinositol**-specific **phospholipase C** (I), purified 447-fold from the culture broth of *B. cereus* by (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> pptn. and chromatog. with CM-Sephadex and DEAE-cellulose, was analyzed in vitro and in vivo for its activity to induce the release of alk. phosphatase from plasma membrane. By i.v. injection of purified I into rats, alk. phosphatase was released into the blood stream quant. Antiserum against I was prepd. and purified to a homogeneous state. Nearly equiv. amts. of purified anti-I IgG completely neutralized both **phosphatidylinositol**-hydrolyzing and phosphatase-releasing activities of the purified I prepn., showing that I is responsible for phosphatase release from rat kidney slices. Also, anti-I IgG inhibited I-induced phosphatase release in vivo. Liberated phosphatase had a mol. wt. of 100,000-110,000 and was derived from organs such as kidney and liver but not from intestine. From in vivo and immunochem. studies, I was demonstrated to be the phosphatasemia factor originally proposed by M. W. Slein and G. F. Logan, Jr. (1965).

L8 ANSWER 32 OF 33 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 6  
ACCESSION NUMBER: 1978:2159 CAPLUS  
DOCUMENT NUMBER: 88:2159  
TITLE: Some characteristics of **phospholipase C** from *Bacillus cereus*  
AUTHOR(S): Otnaess, Anne Brit; Little, Clive; Sletten, Knut;  
Wallin, Reidar; Johnsen, Sven; Flengsrud, Ragnar;  
Prydz, Hans  
CORPORATE SOURCE: Inst. Med. Biol., Univ. Tromsøe, Tromsøe, Norway  
SOURCE: Eur. J. Biochem. (1977), 79(2), 459-68  
CODEN: EJBCAI  
DOCUMENT TYPE: Journal  
LANGUAGE: English

AB The amino acid compn. of purified *B. cereus* **phospholipase C** is reported. The enzyme contains 1 methionine residue; 2 fragments were obtained after CNBr cleavage. The sequence of the N-terminal fragment (25 residues) is reported. Antisera were raised against the enzyme and purified by affinity chromatog. The antisera were monospecific and gave 1 pptn. line with purified as well as with crude **phospholipase C**, showing that no antigenic contaminants were present in the purified preps. used as antigen. The antibodies were purified to the extent that .apprx.2 mols. neutralized 1 enzyme mol. The enzyme was quite resistant to denaturation by urea, Na dodecyl sulfate, or heat (in the presence of 1 mM Zn<sup>2+</sup>). **Phospholipase C** hydrolyzed phosphatidylcholine, phosphatidylethanolamine, and phosphatidylserine. Under the conditions used phosphatidylglycerol, cardiolipin, **phosphatidylinositol**, sphingomyelin, lysophosphatidylcholine, and lysophosphatidylethanolamine were not substrates. Replacement of Zn<sup>2+</sup> by Co<sup>2+</sup> or Ni<sup>2+</sup> or variation of pH (7.2-8.3) did not change the range of substrates. Phosphatidylcholine was the best substrate among the isolated phospholipids and dicaproylphosphatidylcholine was clearly a better substrate than

ACCESSION NUMBER: 1978:144514 BIOSIS  
DOCUMENT NUMBER: BA65:31514  
TITLE: ASYMMETRY OF THE PHOSPHO LIPID BI LAYER OF RAT LIVER  
ENDOPLASMIC RETICULUM.  
AUTHOR(S): HIGGINS J A; DAWSON R M C  
CORPORATE SOURCE: SECT. CYTOL., YALE UNIV. SCH. MED., NEW HAVEN, CONN.  
06510,  
USA.  
SOURCE: BIOCHIM BIOPHYS ACTA, (1977) 470 (3), 342-356.  
CODEN: BBACAO. ISSN: 0006-3002.

FILE SEGMENT: BA; OLD  
LANGUAGE: English

AB The phospholipids of intact microsomal membranes were hydrolyzed 50% by **phospholipase C** of *Clostridium welchii*, without loss of the secretory protein contents of the vesicle, which are therefore not permeable to the phospholipase. Phospholipids extracted from microsomes and dispersed by sonication were hydrolyzed rapidly by **phospholipase C-C.** *welchii* with the exception of **phosphatidylinositol**. Assuming that only the phospholipids of the outside of the bilayer of the microsomal membrane are hydrolyzed in intact

vesicles, the composition of this leaflet was calculated as 84% phosphatidylcholine, 8% phosphatidylethanolamine, 9% sphingomyelin and 4% phosphatidylserine, and that of the inner leaflet 28%

phosphatidylcholine,

37% phosphatidylethanolamine, 6% phosphatidylserine and 5% sphingomyelin. Microsomal vesicles were opened and their contents released in part by incubation with deoxycholate (0.098%) lysophosphatidylcholine (0.005%) or treatment with the French pressure cell. Under these conditions, hydrolysis of the phospholipids by **phospholipase C-C.** *welchii* was increased and this was mainly due to increased hydrolysis of those phospholipids assigned to the inner leaflet of the bilayer, phosphatidylethanolamine and phosphatidylserine. Phospholipase A2 of bee venom and **phospholipase C** of *Bacillus cereus* caused rapid loss of vesicle contents and complete hydrolysis of the membrane phospholipids, with the exception of sphingomyelin which is not hydrolyzed by the former enzyme.

**WEST**[Generate Collection](#)[Print](#)**Search Results - Record(s) 1 through 17 of 17 returned.** 1. Document ID: US 20020034798 A1

L3: Entry 1 of 17

File: PGPB

Mar 21, 2002

PGPUB-DOCUMENT-NUMBER: 20020034798

PGPUB-FILING-TYPE: new

DOCUMENT-IDENTIFIER: US 20020034798 A1

TITLE: HIGH-ACTIVITY PHYTASE COMPOSITIONS

PUBLICATION-DATE: March 21, 2002

## INVENTOR- INFORMATION:

NAME	CITY	STATE	COUNTRY	RULE-47
BARENDESE, RUDOLF CAROLUS MARIA	DELFT		NL	
MEESTERS, GABRIEL MARINUS HENRICUS	DELFT		NL	
ANDELA, CARL SIDONIUS MARIA	DELFT		NL	

US-CL-CURRENT: 435/183; 424/94.1, 424/94.6, 424/94.61, 426/302, 435/195, 435/209,  
435/210[Full](#) [Title](#) [Citation](#) [Front](#) [Review](#) [Classification](#) [Date](#) [Reference](#) [Sequences](#) [Attachments](#) [Claims](#) [KINIC](#) [Drawn Desc](#) [Image](#) 2. Document ID: US 20010046693 A1

L3: Entry 2 of 17

File: PGPB

Nov 29, 2001

PGPUB-DOCUMENT-NUMBER: 20010046693

PGPUB-FILING-TYPE: new

DOCUMENT-IDENTIFIER: US 20010046693 A1

TITLE: Method for improving the activity of enzymes

PUBLICATION-DATE: November 29, 2001

## INVENTOR- INFORMATION:

NAME	CITY	STATE	COUNTRY	RULE-47
Beek, Eddy Van	Geel		BE	
Somers, Ingrid	Turnhout		BE	
Peys, Eric	Balen		BE	
Sas, Benedikt	Oud-Turnhout		BE	

US-CL-CURRENT: 435/183; 426/53, 435/188, 435/195, 435/202, 435/207, 435/212[Full](#) [Title](#) [Citation](#) [Front](#) [Review](#) [Classification](#) [Date](#) [Reference](#) [Sequences](#) [Attachments](#) [Claims](#) [KINIC](#) [Drawn Desc](#) [Image](#) 3. Document ID: US 6500426 B1

L3: Entry 3 of 17

File: USPT

Dec 31, 2002

US-PAT-NO: 6500426

DOCUMENT-IDENTIFIER: US 6500426 B1

TITLE: Carbohydrate-based enzyme-containing granules for use in animal feed

[Full](#) [Title](#) [Citation](#) [Front](#) [Review](#) [Classification](#) [Date](#) [Reference](#) [Sequences](#) [Attachments](#) [Claims](#) [KIMC](#) [Drawn Desc](#) [Image](#) 4. Document ID: US 6183739 B1

L3: Entry 4 of 17

File: USPT

Feb 6, 2001

US-PAT-NO: 6183739

DOCUMENT-IDENTIFIER: US 6183739 B1

TITLE: Phospholipases in animal feed[Full](#) [Title](#) [Citation](#) [Front](#) [Review](#) [Classification](#) [Date](#) [Reference](#) [Sequences](#) [Attachments](#) [Claims](#) [KIMC](#) [Drawn Desc](#) [Image](#) 5. Document ID: US 6143545 A

L3: Entry 5 of 17

File: USPT

Nov 7, 2000

US-PAT-NO: 6143545

DOCUMENT-IDENTIFIER: US 6143545 A

TITLE: Method for reducing phosphorus content of edible oils

[Full](#) [Title](#) [Citation](#) [Front](#) [Review](#) [Classification](#) [Date](#) [Reference](#) [Sequences](#) [Attachments](#) [KIMC](#) [Drawn Desc](#) [Image](#) 6. Document ID: US 6103505 A

L3: Entry 6 of 17

File: USPT

Aug 15, 2000

US-PAT-NO: 6103505

DOCUMENT-IDENTIFIER: US 6103505 A

TITLE: Method for reducing phosphorus content of edible oils

[Full](#) [Title](#) [Citation](#) [Front](#) [Review](#) [Classification](#) [Date](#) [Reference](#) [Sequences](#) [Attachments](#) [KIMC](#) [Drawn Desc](#) [Image](#) 7. Document ID: US 6017530 A

L3: Entry 7 of 17

File: USPT

Jan 25, 2000

US-PAT-NO: 6017530

DOCUMENT-IDENTIFIER: US 6017530 A

TITLE: Phospholipases in animal feed[Full](#) [Title](#) [Citation](#) [Front](#) [Review](#) [Classification](#) [Date](#) [Reference](#) [Sequences](#) [Attachments](#) [KIMC](#) [Drawn Desc](#) [Image](#)

8. Document ID: US 5759537 A

L3: Entry 8 of 17

File: USPT

Jun 2, 1998

US-PAT-NO: 5759537

DOCUMENT-IDENTIFIER: US 5759537 A

TITLE: Animal feeds

[Full](#) [Title](#) [Citation](#) [Front](#) [Review](#) [Classification](#) [Date](#) [Reference](#) [Sequences](#) [Attachments](#)[KMC](#) [Draw Desc](#) [Image](#) 9. Document ID: US 5082674 A

L3: Entry 9 of 17

File: USPT

Jan 21, 1992

US-PAT-NO: 5082674

DOCUMENT-IDENTIFIER: US 5082674 A

TITLE: Food product

[Full](#) [Title](#) [Citation](#) [Front](#) [Review](#) [Classification](#) [Date](#) [Reference](#) [Sequences](#) [Attachments](#)[KMC](#) [Draw Desc](#) [Image](#) 10. Document ID: US 4933192 A

L3: Entry 10 of 17

File: USPT

Jun 12, 1990

US-PAT-NO: 4933192

DOCUMENT-IDENTIFIER: US 4933192 A

TITLE: Hydratable powders which form WOW emulsions

[Full](#) [Title](#) [Citation](#) [Front](#) [Review](#) [Classification](#) [Date](#) [Reference](#) [Sequences](#) [Attachments](#)[KMC](#) [Draw Desc](#) [Image](#) 11. Document ID: WO 9636244 A1

L3: Entry 11 of 17

File: EPAB

Nov 21, 1996

PUB-NO: WO009636244A1

DOCUMENT-IDENTIFIER: WO 9636244 A1

TITLE: APPLICATION OF PHOSPHOLIPASES IN ANIMAL FEED

[Full](#) [Title](#) [Citation](#) [Front](#) [Review](#) [Classification](#) [Date](#) [Reference](#) [Sequences](#) [Attachments](#)[KMC](#) [Draw Desc](#) [Image](#) 12. Document ID: EP 743017 A2

L3: Entry 12 of 17

File: EPAB

Nov 20, 1996

PUB-NO: EP000743017A2

DOCUMENT-IDENTIFIER: EP 743017 A2

TITLE: Application of phospholipases in animal feed

[Full](#) [Title](#) [Citation](#) [Front](#) [Review](#) [Classification](#) [Date](#) [Reference](#) [Sequences](#) [Attachments](#)[KMC](#) [Draw Desc](#) [Image](#)

13. Document ID: WO 200224881 A1 AU 200189588 A

L3: Entry 13 of 17

File: DWPI

Mar 28, 2002

DERWENT-ACC-NO: 2002-383187

DERWENT-WEEK: 200252

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TITLE: New phospholipase from *Zygoascus hellenicus* useful for preparing a dough or baked product, for reducing content of phosphorus in vegetable oil, for production of animal feed and for partial hydrolysis of phospholipids

<a href="#">Full</a>	<a href="#">Title</a>	<a href="#">Citation</a>	<a href="#">Front</a>	<a href="#">Review</a>	<a href="#">Classification</a>	<a href="#">Date</a>	<a href="#">Reference</a>	<a href="#">Sequences</a>	<a href="#">Attachments</a>	<a href="#">KIMC</a>	<a href="#">Drawn Desc</a>	<a href="#">Image</a>
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 14. Document ID: WO 200054604 A1 US 6383485 B1 AU 200037515 A US 6213930 B1 EP 1161153 A1

L3: Entry 14 of 17

File: DWPI

Sep 21, 2000

DERWENT-ACC-NO: 2000-587464

DERWENT-WEEK: 200235

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TITLE: Reducing gastrointestinal inflammation in animals, useful e.g. for improving growth and feeding behavior, comprises reducing release of prostaglandin or leukotriene precursors

<a href="#">Full</a>	<a href="#">Title</a>	<a href="#">Citation</a>	<a href="#">Front</a>	<a href="#">Review</a>	<a href="#">Classification</a>	<a href="#">Date</a>	<a href="#">Reference</a>	<a href="#">Sequences</a>	<a href="#">Attachments</a>	<a href="#">KIMC</a>	<a href="#">Drawn Desc</a>	<a href="#">Image</a>
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 15. Document ID: DE 69716711 E WO 9826057 A1 EP 869167 A2 AU 9851878 A CN 1245532 A JP 2000507458 W US 6103505 A US 6143545 A EP 869167 B1

L3: Entry 15 of 17

File: DWPI

Dec 5, 2002

DERWENT-ACC-NO: 1998-362425

DERWENT-WEEK: 200304

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TITLE: New isolated phospholipase from *Fusarium oxysporum* - used for e.g. reducing phosphorus content of edible oils, treatment of starch hydrolysates, production of animal feed or in detergent or cleaning compositions

<a href="#">Full</a>	<a href="#">Title</a>	<a href="#">Citation</a>	<a href="#">Front</a>	<a href="#">Review</a>	<a href="#">Classification</a>	<a href="#">Date</a>	<a href="#">Reference</a>	<a href="#">Sequences</a>	<a href="#">Attachments</a>	<a href="#">KIMC</a>	<a href="#">Drawn Desc</a>	<a href="#">Image</a>
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 16. Document ID: EP 743017 A2 RO 117142 B1 WO 9636244 A1 AU 9652255 A NL 1003096 C2 ZA 9603861 A CA 2176634 A JP 09098726 A EP 743017 A3 CZ 9700111 A3 BR 9606365 A KR 97704360 A NZ 286574 A AU 700385 B IL 118245 A US 6017530 A US 6183739 B1 CN 1156955 A

L3: Entry 16 of 17

File: DWPI

Nov 20, 1996

DERWENT-ACC-NO: 1996-507458

DERWENT-WEEK: 200225

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**TITLE: Animal feeds contg. phospholipase - used for improving wt. gain and feed efficiency**

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17. Document ID: GB 2267033 A ES 2123644 T3 WO 9422324 A1 AU 9339592 A GB 2267033 B EP 692936 A1 US 5759537 A EP 692936 B1 DE 69319929 E

L3: Entry 17 of 17

File: DWPI

Nov 24, 1993

DERWENT-ACC-NO: 1993-388749

DERWENT-WEEK: 199909

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**TITLE: Increasing growth rate, milk prodn. and quality in livestock - by addn. of phospholipid which increases porosity of rumen or stomach membrane**

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Terms	Documents
L1 same (animal feed)	17

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[Previous Page](#) [Next Page](#)

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L2: Entry 4 of 5

File: USPT

Nov 6, 2001

DOCUMENT-IDENTIFIER: US 6312919 B1

TITLE: Process for producing a cholesterol-reduced substance

Brief Summary Text (337):

As the cholesterol-reduced composition of the present invention, microbial cells or treated materials thereof, crude purified enzymes, purified enzymes and the like containing these three enzymes may be used without any treatment provided they have activities of a cholesterol dehydrogenase, 4-cholest-3-one dehydrogenase and coprostan-3-one dehydrogenase, and further, those in the form of tablets, powders, fine particles, granules, capsules, syrups and the like molded with vehicles which are acceptable for food or medicine may be used. The composition of the present invention may be added as a composition to be added into food or feed for reducing the amount of cholesterol in the food or feed, or may be orally administered via oral route for reducing the cholesterol level in serum. When a crude purified enzyme or purified enzyme is used as the composition of the present invention, the composition optionally may be advantageously prepared so that nicotinamide, phosphate ion or phospholipase is contained in the composition. Examples of the form of the oral composition of the present invention include tablets, powders, fine particles, granules, capsules, syrups, enteric agent, troches and the like. In the case of addition or administration, as the vehicle, any compound such as saccharides like sorbitol, lactose, glucose, lactose, dextrin, starch, crystalline cellulose and the like; inorganic compounds like calcium carbonate, calcium sulfate and the like; distilled water, sesame oil, corn oil, olive oil, cotton seed oil and the like, generally can be used. In preparing the composition, additives such as binder, lubricant, disperser, suspending agent, emulsifying agent, diluent, buffering agent, antioxidant, bacterium inhibiting agent and the like may be used.